





Revised Transcriptome-Based Gene Annotation for Aspergillus flavus Strain NRRL 3357

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ABSTRACT Aspergillus flavus is an agriculturally and medically important filamentous fungus that produces mycotoxins, including aflatoxins, which are potent carcinogens. Here, we generated short- and long-read transcript sequence data from the growth of A. flavus strain NRRL 3357 under both typical and stress conditions to produce a new annotation of its genome.

spergillus flavus (order Eurotiomycetes, phylum Ascomycota) is an agriculturally (1) and medically relevant (2) filamentous fungus that produces diverse secondary metabolites (3). Originally isolated from infected peanut cotyledons in the United States, A. flavus strain NRRL 3357 is available from the ATCC (https://www.atcc.org/). To facilitate the study of the biosynthetic gene clusters involved in secondary metabolism and of the genetic determinants of pathogenicity, we generated a new gene annotation based on transcriptomic sequence data obtained from the growth of strain NRRL 3357 under diverse conditions.

Total RNA was isolated from conidia (asexual spores), mycelia (vegetative cells), and cells undergoing asexual development. Conidia were grown at 30°C on solid glucose minimal medium (GMM) for 2 days prior to RNA extraction. Mycelia were grown at 30°C in liquid GMM, with shaking at 220 rpm for 24 h prior to RNA extraction. Cells undergoing asexual development were grown for 18 h in liquid GMM with shaking at 220 rpm, switched to solid GMM for asexual developmental induction, and grown for an additional 24h at 30°C. To capture mRNA not produced under typical growth conditions, we also isolated RNA from mycelia grown under three separate stress conditions, salinity (0.6 M NaCl), oxidative stress (5 mM H₂O₂), and heat (37°C). After 24 h of normal growth, mycelia were stressed for an additional 24h in GMM at 30°C (or for 24h at 37°C for heat stress). All samples were freeze-dried and ground using a mortar and pestle and then chilled with liquid N₂ to prevent RNA degradation. RNA was isolated from the samples using the Qiagen RNeasy plant minikit and stored at -80° C.

All six samples (RNA from conidia, mycelia, cells undergoing asexual development, and mycelia grown under the three stress conditions) were mixed in equal molar amounts into a single sample for sequencing. We also sequenced the individual samples for mycelia (grown at 30°C in liquid GMM) and heat-stressed mycelia (grown at 37°C in liquid GMM). The Illumina libraries for the three samples were generated using the Lexogen mRNA Sense v2 library preparation kit (Illumina) and were sequenced on the HiSeq X Ten platform (2×150 -bp paired-end format) by Novogene, Ltd. (Beijing). Citation Hatmaker EA, Zhou X, Mead ME, Moon H. Yu. J-H. Rokas A. 2020. Revised transcriptome-based gene annotation for Aspergillus flavus strain NRRL 3357. Microbiol Resour Announc 9:e01155-20. https://doi.org/ 10.1128/MRA.01155-20.

Editor Jason E. Stajich, University of California,

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Received 1 October 2020 Accepted 15 November 2020 Published 3 December 2020

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TABLE 1 Summary of BUSCO assessments of the Aspergillus flavus NRRL 3357 genome assembly annotations

	No. (%) of genes determined by BUSCO		
Type of data	Complete	Fragmented	Missing
Annotation (NCBI) ^a	3,559 (88.0)	400 (9.9)	87 (2.1)
Annotation (AspGD) ^b	3,659 (90.5)	261 (6.5)	126 (3.0)
Annotation (this study)	3,896 (96.3)	106 (2.6)	44 (1.1)

^a See ftp://ftp.ncbi.nlm.nih.gov/genomes/all/GCF/000/006/275/GCF_000006275.2_JCVI-afl1-v2.0/GCF 000006275.2 JCVI-afl1-v2.0_protein.faa.gz.

Totals of 27,714,710 (library from mix of six samples), 29,581,787 (mycelia grown at 30° C), and 24,719,957 (mycelia grown at 37°C) raw reads were generated. An additional Iso-Seq library was prepared from the mix of the six samples, size selected for fragments of ≥4 kb using the BluePippin size selection system (Sage Science), and analyzed on the RS II instrument (Pacific Biosciences), resulting in 5,173,949 reads with a mean length of 2,785 bp. Short and long reads were used together to generate a de novo transcriptome assembly using Trinity v.2.10.0 (4). As part of the Trinity assembly pipeline, low-quality bases and adaptor contamination were trimmed from the Illumina reads using Trimmomatic v.0.39.0 (5). Short reads were also used for genome-guided assembly using Trinity.

Initial gene models were generated using default parameters for MAKER v.2.31.9 (6), which included three ab initio gene predictors, AUGUSTUS v.3.3.3 (7), GeneMark-ES v.4.39 (8), and SNAP v.2013-11-29 (9). Exonerate v.2.4.0 (10) was provided for alignment to the MAKER pipeline. Isoforms were added to PASA v.2.3.3 (11) from both the de novo and genome-quided transcriptome assemblies as "EST evidence." The new annotation predicts 14,313 genes and 14,699 proteins. We compared our annotation to the Aspergillus flavus NRRL 3357 genome annotations from AspGD and NCBI (accession number GCA_000006275.2) using BUSCO v.3.0.2 (12), which estimated a copy number of 4,046 preselected genes considered single copy within the OrthoDB (13) database (Table 1). The new annotation contains 96.3% of these 4,046 genes, substantially higher than the percentages contained by the previous two annotations (AspGD, 90.5%; NCBI, 88.0%; Table 1). The transcriptome-based annotation contains 12,569 genes which overlap one or more gene models in the previous NCBI annotation, as well as 1,744 predicted genes without any overlap with the previous annotation, suggesting that our transcriptome-based annotation provides new information for studying A. flavus.

Data availability. A. flavus NRRL 3357 RNA sequences from this study have been deposited under Sequence Read Archive (SRA) study SRP213830, with individual samples deposited in the SRA under accession numbers SRX6419472, SRX6419473, SRX6419474, and SRX6419475 and BioSample accession numbers SAMN12235769 through SAMN12235771. This whole-genome sequencing project has been deposited at DDBJ/ENA/GenBank under the accession number AAIH00000000. The annotation has been updated for A. flavus NRRL 3557 assembly accession number GCA_000006275.3 (GenBank), and the annotation described in this paper is the third version, AAIH03000000.

ACKNOWLEDGMENT

We thank William C. Nierman for giving us permission to update the gene annotation of Aspergillus flavus NRRL 3357.

REFERENCES

- 1. Klich MA. 2007. Aspergillus flavus: the major producer of aflatoxin. Mol Plant Pathol 8:713-722. https://doi.org/10.1111/j.1364-3703 .2007.00436.x.
- 2. Cunha C, Aversa F, Romani L, Carvalho A. 2013. Human genetic
- susceptibility to invasive aspergillosis. PLoS Pathog 9:e1003434. https:// doi.org/10.1371/journal.ppat.1003434.
- 3. Amaike S, Keller NP. 2011. Aspergillus flavus. Annu Rev Phytopathol 49:107-133. https://doi.org/10.1146/annurev-phyto-072910-095221.

^b See http://www.aspergillusgenome.org/download/sequence/A_flavus_NRRL_3357/current/A_flavus_NRRL 3357 orf trans all.fasta.gz.

Downloaded from https://journals.asm.org/journal/mra on 17 February 2022 by 210.59.240.222.

- 4. Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, Adiconis X, Fan L, Raychowdhury R, Zeng Q, Chen Z, Mauceli E, Hacohen N, Gnirke A, Rhind N, di Palma F, Birren BW, Nusbaum C, Lindblad-Toh K, Friedman N, Regev A. 2011. Trinity: reconstructing a full-length transcriptome without a genome from RNA-Seq data. Nat Biotechnol 29:644.
- 5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120.
- 6. Holt C, Yandell M. 2011. MAKER2: an annotation pipeline and genome-database management tool for second-generation genome projects. BMC Bioinformatics 12:491.
- 7. Stanke M, Morgenstern B. 2005. AUGUSTUS: a Web server for gene prediction in eukaryotes that allows user-defined constraints. Nucleic Acids Res 33:W465-W467.
- 8. Ter-Hovhannisyan V, Lomsadze A, Chernoff YO, Borodovsky M. 2008. Gene prediction in novel fungal genomes using an ab initio algorithm with unsupervised training. Genome Res 18:1979–1990.

- 9. Korf I. 2004. Gene finding in novel genomes. BMC Bioinformatics 5:59.
- 10. Slater GSC, Birney E. 2005. Automated generation of heuristics for biological sequence comparison. BMC Bioinformatics 6:31.
- 11. Haas BJ, Salzberg SL, Zhu W, Pertea M, Allen JE, Orvis J, White O, Robin CR, Wortman JR. 2008. Automated eukaryotic gene structure annotation using EVidenceModeler and the Program to Assemble Spliced Alignments. Genome Biol 9:R7.
- 12. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics 31:3210-3212.
- 13. Zdobnov EM, Tegenfeldt F, Kuznetsov D, Waterhouse RM, Simao FA, Ioannidis P, Seppey M, Loetscher A, Kriventseva EV. 2017. OrthoDB v9.1: cataloging evolutionary and functional annotations for animal, fungal, plant, archaeal, bacterial and viral orthologs. Nucleic Acids Res 45:D744-D749.